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509.BONE MARROW FAILURE AND CANCER PREDISPOSITION SYNDROMES: CONGENITAL

Hnrnpk Overexpression Drives Nucleolar Aberrancies Causing Ribosomopathies

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Background: Protein biogenesis is a complex process involving *nucleoli* and ribosomes. Alterations in any step could lead to alterations in ribosome functionality and protein synthesis. Hnrnpk is an RNA-binding protein (RBP) involve in these processes, finding that an overexpression (OE) produces nucleus and nucleolar stress (NS), decreases transcription, and drives an imbalance in ribosome biogenesis, causing a reduced translation.

Aims: To elucidate how hnRNP K dysregulation affects the hematopoietic stem cell (HSCs) biology.

Methods: To study the impact of Hnrnpk OE *in vivo*, we developed an inducible tamoxifen mouse model, *Hnrnpk*^{Tg/hUbc-CreERT2}. Survival was evaluated by Kaplan-Meier, phenotype was described by symptoms/signs, CBC, bone marrow (BM) H/E, IHC and FCM analysis, and serum IL-6 ELISA.

HSCs were cultured to study the impact of Hnrnpk OE in the HSCs dynamics. Hnrnpk OE was established *in vitro* using CRISPR/SAM. RNA-seq analysis was performed in a single read 85-base format and analyzed with DESeq2. TMT-based deep proteome profiling was also performed. Both were GSEA preranked.

Transcription and translation were tested using Click-it RNA and HPG kit respectively, and translation efficiency by polysome assay. NS were analyzed by confocal microscopy and transmission electron microscopy (TEM). Protein-protein interaction between Hnrnpk and Ncl was studied by IP. Possible phenotype rescue was carried using *Hnrnpk*^{Tg/hUbc-CreERT2}/ *c-Myc*^{lox/wt}, *Hnrnpk*^{Tg/hUbc-CreERT2}/ Tp53^{lox/wt} and *Hnrnpk*^{Tg/hUBC-CreERT2}/ *Ncl*^{KD} *in vitro* and *in vivo* models. Cell cycle FACS, senescence assays and karyotyping were performed. Molecular mechanism was elucidated by qRT-PCR and WB.

Results: Hnrnpk ^{Tg/hUbc-CreERT2} mice had widespread Hnrnpk OE and lifespan's reduction. By CBC, we found the development of leukopenia, lymphopenia, anaemia and thrombocytopenia (Fig.A). BM H/E, IHC and FCM showed a reduction of B220 ⁺ and CD34 ⁺ and Sca1 ⁺ HSCs, and an increment in myeloid cells (Fig. B). Also, we found higher senescent β -galactosidase expression in BM and IL-6 *in vivo* (Fig.C). Then, we found a decay in viability and an exhaustion in HSCs (Fig.D).

To understand Hnrnpk implication in BM failure phenotype *in vivo*, we generated Hnrnpk OE cells (Fig.E). RNA-seq showed an upregulation in G2/M-checkpoint pathway related molecules (Fig.F), confirmed by FACS analysis, showing and increment of arrested G2/M phase-cells (Fig.G). Moreover, we showed a rise in β -galactosidase activity, polyploidy and genomic instability

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Session 509

(Fig. H), linked to an increment in p21 and p16 (Fig.I). Then, TEM revealed nucleolar alterations in the *in vitro* model, including segregation, anormal accumulations or fragmentation of nucleolar components, alterations validated by confocal microscopy (Fig.J-K). Also, we found a Ncl increment *in vitro*, consistent with the protein-protein interaction between Hnrnpk and Ncl.

Then, proteomics showed that Hnrnpk OE correlates with ribosome biogenesis regulators, c-Myc and mTOR dysregulation (Fig.L). Moreover, we found a decrease in transcription (Fig.M), consistent with rRNAs reduction, driving translation and protein synthesis deficiency (Fig.N). There was a partial reversion of NS hallmarks in *Hnrnpk* ^{Tg/hUbc-CreERT2}/ *c-Myc* ^{lox/wt}, *Hnrnpk* ^{Tg/hUbc-CreERT2}/ *Tp53* ^{lox/wt} and *Hnrnpk* ^{Tg/hUBC-CreERT2}/ *Ncl* ^{KD} MEFs *in vitro* models and partially rescue phenotype *in vivo* (Fig.O). Finally, we focused on the nucleolus. Thus, Hnrnpk OE cells showed a reduction in Fbl, increase in Ncl and Ncl diffusion in nucleoplasm. All these data suggest the existence of a ribosompathy-like phenotype.

Conclusions: this work found that RBPs dysregulation such as Hnrnpk OEdrives BM failure phenotype, promoting the exhaustion of HSCs by nucleoulus/ribosome dysregulation that triggers cell cycle arrest and apoptosis, dependent of p53. We therefore suggest that Hnrnpk induces ribosome dysfunction consistent with some types of ribosomopathies.

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Figure A: H&E and hnRNP K IHC in mice BM; WB showing hnRNP K in liver and spleen of *in vivo* model; CBC: total WBC (p=0.032), % lymphocytes (p=0.001), % myeloid cells (p=0.017) and % platelets (p=0.009), total red blood cells (p=0.016), hematocrit (p=0.050) and hemoblobin (p=0.017) **. Figure B**: H&E and B220, Gr1 and Mpo IHC in mice BM; FCM of total BM, Cd11b*/Gr-1*(p = 0.0327*); H&E and Cd34 IHC in mice BM; FCM of total BM, Cd34* (p = 0.0208*) and Sca-1* (p = 0.0424*). Figure C: H&E and β-galactosidase IHC in mice BM; IFCM of ELISA from Hnrnpk^{TighUBC-CHERT} mice (+4-OHT) vs Hnrnpk^{Tig} (+4-OHT) vs Hnrnpk^{Tig} (+4-OHT) vs Hnrnpk^{TighUBC-CHERT} mice (+4-OHT) vs Hnrnpk^{Ti}

